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(54) Title: BIOCIDAL PROTEINS (57) Abstract Biocidal proteins isolated from <i>Amaranthus</i> have been characterised. The proteins show a wide range of antifungal activity and are active against gram-positive bacteria. DNA encoding the proteins has been isolated and incorporated into vectors. Plants may be transformed with this DNA. The proteins find commercial application as antifungal or antibacterial agents; transgenic plants will show increased disease resistance.		

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BIOCIDAL PROTEINS

This invention relates to biocidal proteins, processes for their manufacture and use, and DNA sequences coding for them. In particular, it relates to antimicrobial proteins isolated from Amaranthus.

Amaranthus caudatus (amaranth) belongs to a large family, the Amaranthaceae, of herbs and shrubs which grow widely in tropical, sub-tropical and temperate regions. Amaranth is an ancient food crop of the Americas, and is still cultivated for grain production in parts of Central and South America, Asia and Africa. Amaranth seeds can be popped, toasted, cooked for gruel, milled into flour or made into flat breads, and have a particularly high nutritive value (Betschart et al, 1981, J Food Sci, 46, 1181-1187; Pedersen et al, 1987, Plant Foods Hum Nutr, 36, 309-324). Amaranth is also cultivated world-wide as a garden ornamental.

Although plants normally grow on substrates that are extremely rich in fungal organisms, infection remains a rare event. To keep out potential invaders, plants produce a wide array of antifungal compounds, either in a constitutive or an inducible manner. The best studied of these are phytoalexins, secondary metabolites with a broad antimicrobial activity spectrum that are specifically synthesised upon perception of appropriate defence-related signal molecules. The production of phytoalexins depends on the transcriptional activation of a series of genes encoding enzymes of the phytoalexin biosynthetic

pathway. During the last decade, however, it has become increasingly clear that some plant proteins can play a more direct role in the control of phytopathogenic fungi. Several classes of proteins with antifungal properties have now been identified, including beta-1,3-glucanases, ribosome-inactivating proteins, thionins, chitinases, chitin-binding lectins and zeamatin.

Chitin (poly- β -1,4-N-acetyl-D-glucosamine) is a polysaccharide occurring in the cell wall of fungi and in the exoskeleton of invertebrates. Although plants do not contain chitin or chitin-like structures, proteins exhibiting strong affinity to this polysaccharide have been isolated from different plant sources (Raikhel and Broekaert, 1991, in: Verma, ed, Control of plant gene expression, in press).

Examples of such chitin-binding proteins are basic chitinases from bean (Boller et al, 1983, Planta, 157:22-31), wheat (Molano et al, 1979, J Biol Chem, 254:4901-4907) and tobacco (Shinshi et al, 1987, Proc Natl Acad Sci USA, 84:89-93); chitin-binding lectins from wheat (Rice and Etzler, 1974, Biochem Biophys Res Comm, 59:414-419), barley (Peumans et al, 1982, Biochem J, 203:239-143), rice (Tsuda, 1979, J Biochem, 86:1451-1461), and stinging nettle (Peumans, et al, 1983, FEBS Lets, 177:99-103); and a small protein from rubber tree latex called hevein (Van Parijs et al, 1991, Planta, 183:258-264). All these chitin-binding proteins share a homologous cysteine/glycine-rich domain of about 40-43 amino acids, which is either repeated two-fold (in the nettle lectin), four-fold (in wheat, barley and rice lectins) or fused to an

unrelated domain (in basic chitinases).

The exact physiological role of these proteins remains uncertain, but they all show antibiotic activity in vitro, suggesting a defence-related function. Antifungal properties have been ascribed to chitinases (Schlumbaum et al, 1986, Nature, 324:365-367; Broekaert et al, 1988, Physiol Mol Plant Pathol, 33:319-331), nettle lectin (Broekaert et al, 1989, Science, 245:1100-1102) and hevein (Van Parijs et al, 1991, Planta, 183:258-264). The wheat lectin causes deleterious effects on the development of insect larvae (Murdock et al, 1990, Phytochem, 29:85-89; Czapla and Lang, 1990, J Econ Entomol, 83:2480-2485). However, it is not established whether or not the observed antibiotic effects on fungi or insects are related to the chitin-binding activity of these proteins.

Successful application of chitin-binding proteins, especially chitinases, in the protection of plants against fungal disease has been reported. In US Patent Number 4940840 (DNA Plant Technology Corporation), tobacco plants expressing a chitinase gene from the bacterium Serratia marcescens appear to be less sensitive to the fungus Alternaria longipes. European Patent Application Number 418695 (Ciba Geigy) describes the use of regulatory DNA sequences from tobacco chitinase gene to drive expression of introduced genes producing transgenic plants with improved resistance to pathogens. Patent Application Number WO9007001 (Du Pont de Nemours Company) describes production of transgenic plants which over-express a chitinase gene giving improved resistance to fungal pathogens.

We have now purified a new class of potent antimicrobial proteins which exhibit chitin-binding properties.

5 According to the present invention, we provide antimicrobial proteins capable of being isolated from seeds of Amaranthus.

10 In further aspects, this invention comprises a vector containing a DNA sequence coding for a protein according to the invention. The DNA may be cloned or transformed into a biological system allowing expression of the encoded protein.

The invention also comprises plants transformed with recombinant DNA encoding an antimicrobial protein according to the invention.

15 The invention also comprises a process of combatting fungi or bacteria whereby they are exposed to the proteins according to the invention.

A new class of potent antimicrobial proteins has been isolated from seeds of Amaranthus caudatus (amaranth). The class includes two protein factors, hereafter called Ac-AMP1 (Amaranthus caudatus - Antimicrobial Protein 1) and Ac-AMP2 (Amaranthus caudatus - Antimicrobial Protein 2) respectively. Both are dimeric proteins, composed of two identical 3 kDa subunits. Both proteins are highly basic and have pI values above 10. Proteins with similar antifungal activity have been extracted from the seed of several closely related species, including Amaranthus paniculatus, Amaranthus retroflexus, Amaranthus lividus and Gomphrena globosa.
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The amino acid sequence of Ac-AMP1 (29 residues) is identical to that of Ac-AMP2 (30 residues), except that the latter has one

additional residue at the carboxyl- terminus.
Knowledge of these sequences enables manufacture of
the Ac-AMP proteins using a standard peptide
synthesiser.

5 The amino acid sequences of Ac-AMP1 and
Ac-AMP2 are highly homologous to the
cysteine/glycine-rich domain occurring in many
chitin-binding proteins. Moreover, Ac-AMP1 and
Ac-AMP2 bind to chitin and can be desorbed at low
10 pH (a property shared by chitinases and lectins),
and so can be considered as new members of the
family of chitin-binding proteins. However, when
compared to the regular 40-43 amino acid
cysteine/glycine-rich domains found in the
15 chitin-binding proteins previously characterised,
the Ac-AMPs distinguish themselves by several
features. These include a higher abundance of
basic amino acids, the presence of an additional
amino-terminal residue, the occurrence of a gap of
20 four amino acids at position 6 to 9, and the lack
of a carboxyl-terminal portion of 10-12 residues.

 cDNA encoding the Ac-AMPs has been isolated
and sequenced. The cDNA encoding Ac-AMP2 has been
identified. It encodes an 86-amino acid preprotein
25 and a 25-amino acid carboxy-terminal extension.
The structure of this preprotein differs from all
other known precursors of chitin-binding proteins.
The cDNA encoding Ac-AMP1 has been identified as a
post-translational cleavage product of Ac-AMP2.

30 This DNA can be manufactured using a standard
nucleic acid synthesiser, or suitable probes
(derived from the known sequence) can be used to
isolate the actual Ac-AMP gene(s) and control
sequences from the plant genome. This genetic

material can then be cloned into a biological system which allows expression of the proteins under the control of a constitutive or inducible promoter. Hence the proteins can be produced in a suitable micro-organism or cultured cell, extracted and isolated for use. Suitable micro-organisms include Escherichia coli and Pseudomonas. Suitable cells include cultured insect cells and cultured mammalian cells. The DNA can also be transformed by known methods into any plant species, so that the antimicrobial proteins are expressed within the plant.

Plant cells according to the invention may be transformed with constructs of the invention according to a variety of known methods (Agrobacterium Ti plasmids, electroporation, microinjection, microprojectile gun, etc). The transformed cells may then in suitable cases be regenerated into whole plants in which the new nuclear material is stably incorporated into the genome. Both transformed monocot and dicot plants may be obtained in this way, although the latter are usually more easy to regenerate.

Examples of genetically modified plants according to the present invention include: fruit such as tomatoes, mangoes, peaches, apples, pears, strawberries, bananas and melons; field crops such as canola, sunflower, tobacco, sugarbeet, small-grain cereals such as wheat, barley and rice, maize and cotton: and vegetables such as carrot, lettuce, cabbage and onion.

The Ac-AMP proteins show a wide range of antifungal activity, and are also active against gram-positive bacteria. The proteins could be used

as fungicides or antibiotics by application to plant parts. The proteins could also be used to combat fungal or bacterial disease by expression within plant bodies.

5 Both Ac-AMP1 and Ac-AMP2 show surprisingly high activity: they inhibit the growth of a variety of plant pathogenic fungi at much lower doses than previously-known antifungal chitin-binding proteins. The antifungal effect of the novel
10 proteins is antagonised by Ca^{2+} .

Some chitin-binding proteins are known to have an effect against insects which possess an exoskeleton comprising chitin. The sequence similarity between the Ac-AMPs and known
15 chitin-binding proteins implies that the Ac-AMPs may also possess insecticidal properties.

The Ac-AMP proteins can be isolated and purified from Amaranthus caudatus seeds, synthesised artificially from their known amino
20 acid sequence, or produced within a suitable micro-organism by expression of recombinant DNA. The proteins may also be expressed within a transgenic plant.

The invention may be further understood by reference to the drawings, in which:

Figure 1 shows the cation exchange chromatogram for the antifungal proteins and the associated graph of fungal growth inhibition.

Figure 2A shows the HPLC profile of purified
30 Ac-AMP1.

Figure 2B shows the HPLC profile of purified Ac-AMP2.

Figure 3 shows the SDS-PAGE analysis of the purified antifungal proteins.

Figure 4A shows the amino acid sequences of Ac-AMP1 and Ac-AMP2.

Figure 4B shows the alignment of amino acid sequences from tobacco chitinase, bean chitinase, 5 hevein, wheat lectin, nettle lectin, and Ac-AMP2.

Figure 5 shows the SDS-PAGE analysis of different eluates after affinity chromatography of antifungal proteins on chitin.

Figure 6 shows the growth inhibition curves of 10 fungi measured at varying concentrations of antifungal proteins.

Figure 7 shows the growth inhibition curves of B cinerea measured at varying concentrations of antifungal proteins with and without different 15 additions of KCl or CaCl₂.

Figure 8 shows the nucleotide sequence and deduced amino acid sequence of a cDNA clone encoding Ac-AMP2.

Figure 9 shows the structure of the expression 20 vectors pAC11 and pAC12.

Figure 10 shows the structure of the plant transformation vectors pAC111 and pAC112.

The following Examples illustrate the invention.

25

EXAMPLE 1

Extraction of basic heat-stable proteins from Amaranthus caudatus seeds.

Ammonium sulphate fractionation of proteins 30 precipitating in the interval of 30 to 75% relative saturation was followed by isolation of the basic protein fraction (pI>9) by passage over a Q-Sepharose (Pharmacia) anion exchange column equilibrated at pH 9. The detailed methods are

described below.

One kg of A caudatus seeds (obtained from Gonthier, Wanze, Belgium) was ground in a coffee mill and the resulting meal was extracted for 2
5 hours at 4°C with 3 litres of an ice-cold extraction buffer containing 10 mM NaH_2PO_4 , 15 mM Na_2HPO_4 , 100 mM KCl, 2 mM EDTA, 2 mM thiourea, 1 mM PMSF and 1 mg/l leupeptin. The homogenate was squeezed through cheesecloth and clarified by
10 centrifugation (5 min at 7,000 x g). Solid ammonium sulphate was added to the supernatant to obtain 30% relative saturation and the precipitate formed after standing for 1 hour at room temperature was removed by centrifugation (10 min
15 at 7,000 x g). The supernatant was adjusted to 75% relative ammonium sulphate saturation and the precipitate formed overnight at room temperature collected by centrifugation (30 min at 7,000 x g). After redissolving the pellet in 300 ml distilled
20 water, the insoluble material was removed by further centrifugation (20 min at 7,000 x g). The clear supernatant was dialyzed extensively against distilled water using benzoylated cellulose tubing (Sigma, St Louis, MO) with a molecular weight cut
25 off of 2,000 Da. After dialysis the solution was adjusted to 50 mM Tris-HCl (pH 9) by addition of the ten-fold concentrated buffer, and subsequently passed over a Q-Sepharose Fast Flow (Pharmacia, Uppsala, Sweden) column (12 x 5 cm) in equilibrium
30 with 50 mM Tris-HCl (pH 9). The proteins passed through the column were dialyzed extensively against 20 mM sodium phosphate buffer (pH 7).

This material represents the basic protein fraction of A caudatus seeds. Its further

chromatographic purification is described in Example 3.

EXAMPLE 2

5 Antifungal and antibacterial activity assays.

Antifungal activity was measured by
microspectrophotometry as previously described
(Broekaert, 1990, FEMS Microbiol Lett, 69:55-60).
Routinely, tests were performed with 20 μ l of a
10 (filter-sterilized) test solution and 80 μ l of a
fungal spore suspension (2×10^4 spores/ml) or
mycelium fragments in half strength Potato Dextrose
Broth (Difco). For experiments on the antagonistic
effect of cations, a synthetic growth medium was
15 used instead of Potato Dextrose Broth. The
synthetic growth medium consisted of K_2HPO_4 (2.5
mM), $MgSO_4$ (50 μ M), $CaCl_2$ (50 μ M), $FeSO_4$ (5 μ M),
 $CoCl_2$ (0.1 μ M), $CuSO_4$ (0.1 μ M), Na_2MoO_4 (2 μ M),
 H_3BO_3 (0.5 μ M), KI (0.1 μ M), $ZnSO_4$ (0.5 μ M), $MnSO_4$
20 (0.1 μ M), glucose (10 g/l), asparagine (1 g/l),
methionine (20 mg/l), myo-inositol (2 mg/l), biotin
(0.2 mg/l), thiamine-HCl (1 mg/l), and
pyridoxine-HCl (0.2 mg/l). Control microcultures
contained 20 μ l of sterile distilled water and 80
25 μ l of the fungal spore suspension.

Unless otherwise stated the test organism was
Fusarium culmorum (strain IMI 180420) and
incubation was carried out at 25°C for 48 hours.
Percent growth inhibition is defined as 100 times
30 the ratio of the corrected absorbance of the
control microculture minus the corrected absorbance
of the test microculture over the corrected
absorbance at 595 nm of the control microculture.
The corrected absorbance values equal the

absorbance at 595 nm of the culture measured after 48 hours minus the absorbance at 595 nm measured after 30 min. Values of growth inhibition lower than 15% are not indicated on the chromatograms.

5 The antifungal activity (units per ml) is calculated as 50 times the dilution factor of a test solution at which 50% growth inhibition is obtained under the given assay conditions.

Antibacterial activity was measured
10 microspectrophotometrically as follows. A bacterial suspension was prepared by inoculating soft nutrient agarose (tryptone, 10 g/l; Seaplaque agarose (FMC), 5 g/l) and kept at 37°C to prevent solidification. Aliquots (80 μ l) of the bacterial
15 suspension (10^5 colony forming units per ml) were added to filter-sterilized samples (20 μ l) in flat-bottom 96-well microplates. The absorbance at 595 nm of the culture was measured with the aid of a microplate reader after 30 minutes and 24 hours
20 of incubation at 28°C. Percent growth inhibition was calculated as described above for the antifungal activity assay.

EXAMPLE 3

25 Purification of antifungal proteins from A caudatus seeds.

The starting material for the isolation of the A caudatus antifungal proteins was the basic protein fraction extracted from the mature seeds as
30 in Example 1. These proteins were further separated by cation exchange chromatography, as shown in Figure 1.

About 100 mg of the basic protein fraction dissolved in 20 mM sodium phosphate buffer (pH 7)

was applied on a S-Sepharose High Performance (Pharmacia) column (10 x 1.6 cm) previously equilibrated with the sodium phosphate buffer. The column was eluted at 3 ml/min with a linear
5 gradient of 210 ml from 0 to 150 mM NaCl in 20 mM sodium phosphate buffer (pH 7). The eluate was monitored for protein by online measurement of the absorbance at 280 nm (results shown in the lower panel of Figure 1) and collected in 7.5 ml
10 fractions of which 20 μ l was tested in the microspectrophotometric antifungal activity assay described in Example 2 (results shown in the upper panel of Figure 1).

Upon fractionation, the mixture resolved into
15 four distinct peaks (Figure 1). The antifungal activity co-eluted with the material from peaks 2 and 4, respectively.

The active fractions were finally purified by reversed-phase chromatography. About 1 mg amounts
20 of peak 2 material (Figure 2A) and peak 4 material (Figure 2B) were loaded on a Pep-S (porous silica, C₂/C₁₈, Pharmacia) column (25 x 0.93 cm) in equilibrium with 0.1% TFA. The column was eluted at 5 ml/min with the following gradients (solvent B is methanol containing 0.1 % TFA): 0-3 min, 0-15%
25 B; 3-23 min, 15-35% B; 23-25 min, 35-100% B. The eluate was monitored for protein by online measurement of the absorption at 280 nm. Five ml fractions of the eluate were collected,
30 vacuum-dried, and finally dissolved in 0.5 ml distilled water of which 10 μ l was used in a microspectrophotometric antifungal activity assay.

Figure 2A and Figure 2B show the HPLC profiles of purified peak 2 and peak 4 material

respectively. The lower panels show monitoring of the eluate for protein by measurement of the absorption at 280 nm. Results of the microspectrophotometric antifungal activity assay are shown in the upper panels.

Both material from peak 2 and from peak 4 yielded well resolved major peaks that co-eluted with the antifungal activity. The active factor purified from peak 2 is called Ac-AMP1 (Amaranthus caudatus antifungal protein 1), and that from peak 4 is designated analogously as Ac-AMP2.

EXAMPLE 4

Molecular structure of the purified antifungal proteins.

The molecular structure of the Ac-AMPs was further analysed. Sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) was performed on precast commercial gels (PhastGel High Density from Pharmacia) using a PhastSystem (Pharmacia) electrophoresis apparatus. The sample buffer contained 200 mM Tris-HCl (pH 8.3), 1% (w/v) SDS, 1 mM EDTA, 0.005% bromophenol blue and, unless otherwise stated, 1% (w/v) dithiothreitol (DTT). Proteins were fixed after electrophoresis in 6% glutaraldehyde and silver-stained according to Heukeshoven and Dernick (1985, Electrophoresis, 6:103-112).

The amaranth antifungal proteins were analysed by SDS-PAGE before and after reduction with dithiothreitol. Reduced Ac-AMP1 and Ac-AMP2 both migrated as single bands with an apparent molecular weight of about 3 kDa. However, in their unreduced state, Ac-AMP1 and Ac-AMP2 yielded a 4 kDa and a 6

kDa band respectively. It appears therefore that the antifungal factors are dimeric proteins stabilised by disulphide bridges, each comprised of two identical 3 kDa subunits. Attempts to
5 determine the molecular weight of the native Ac-AMPs by gel filtration on either Superose-12 or Superdex-75 (Pharmacia) were unsuccessful as the proteins were retarded.

Figure 3 shows the SDS-PAGE analysis of the
10 purified antifungal proteins: lane 1 is reduced Ac-AMP1, lane 2 is reduced Ac-AMP2, lane 3 is unreduced Ac-AMP1, lane 4 is unreduced Ac-AMP2. Two hundred nanograms of the proteins were separated on the gels. Lane M shows myoglobin
15 fragments used as molecular weight markers (Pharmacia) with the following sizes: 17 kDa, 14.5 kDa, 8 kDa, 6 kDa, and 2.5 kDa.

Free cysteine thiol groups were assessed qualitatively as follows. Hundred μ g amounts of
20 reduced or unreduced proteins were dissolved in 6 M guanidinium-Cl containing 100 mM sodium phosphate buffer (pH 7) and 1 mM EDTA. The mixtures were allowed to react with 5,5'-dithionitrobenzoic acid and monitored for release of nitrothiobenzoate as
25 described by Creighton (1989, Protein structure, a practical approach, 155-167). Reduction of the proteins was done by addition of Tris-HCl (pH 8.6) to 100 mM and dithiothreitol to 30 mM, followed by incubation at 45°C for 1 hour. The proteins were
30 separated from the excess reagents by reversed-phase chromatography on a C₂/C₁₈ silica column.

The unreduced Ac-AMPs did not contain free cysteine thiol groups, whereas the reduced proteins

did, indicating that all cysteine residues participate in disulphide bonds. The presence of a relatively high number of disulphide linkages in such small polypeptides suggests that the Ac-AMPs have compact structures.

The pI values of Ac-AMP1 and Ac-AMP2 were determined by isoelectric focusing and found to be 10.3 and over 10.6 respectively. Isoelectric focusing was performed on precast Immobiline Dry Strips (Pharmacia) rehydrated in 8 M urea, using marker proteins in the pI range from 4.7 to 10.6 (Pharmacia).

EXAMPLE 5

Amino acid sequencing of the Ac-AMPs.

Cysteine residues of the antifungal proteins were modified by S-carboxyamidomethylation as follows: 100 μ g amounts of purified proteins were dissolved in 150 μ l 0.3 M Tris-HCl (pH 8.6) containing 30 mM DTT and reacted for 1 hour at 45°C. Iodoacetamide was added to a final concentration of 100 mM and the mixture was kept in the dark at 37°C for 1 hour. The reaction was finally quenched by addition of DTT to a final concentration of 100 mM and allowed to react for an additional hour at 37°C. Removal of excess reagents was done by reversed-phase chromatography. The resulting protein fractions were subjected to amino acid sequence analysis in a 477A Protein Sequencer (Applied Biosystems) with on-line detection of phenylthiohydantoin amino acid derivatives in a 120A Analyser (Applied Biosystems).

The amino acid sequence of the reduced and

carboxyamidomethylated antifungal proteins was determined by direct N-terminal sequencing. Figure 4A shows the N-terminal amino acid sequences of Ac-AMP1 and Ac-AMP2. Ac-AMP1 is 29 amino acids in length, whereas Ac-AMP2 has 30 residues. The sequence of Ac-AMP2 is identical to that of Ac-AMP1 except that it has one additional amino acid at its carboxyl terminus (arginine). The Ac-AMPs are particularly rich in cysteine (6 residues), glycine (6 residues) and basic amino acids (4 and 5 residues for Ac-AMP1 and Ac-AMP2 respectively).

Ac-AMP1 appears to be a truncated form of Ac-AMP2. It is possible that the two proteins result from the same precursor molecule by differential post-translational processing.

The theoretical isoelectric points calculated from the sequence data are 10.1 and 11.0 for Ac-AMP1 and Ac-AMP2 respectively, assuming that all cysteine residues participate in disulphide linkages. These compare well to the measured pI values given in Example 4.

Figure 4B shows the alignment of N-terminal amino acid sequences from tobacco chitinase (Shinshi et al, 1987, Proc Natl Acad Sci USA, 84:89-93), bean chitinase (Broglie et al, 1986, Proc Natl Acad Sci USA, 83:6820-6824), hevein (Broekaert et al, 1990, Proc Natl Acad Sci USA, 87:7633-7637), wheat lectin (Raikhel and Wilkins, 1987, Proc Natl Acad Sci USA, 84:6745-6749), nettle lectin (Chapot et al, 1986, FEBS Lett, 195:231-234) and the sequence of Ac-AMP2. Sequence identities with the tobacco chitinase are indicated in capitals, conserved changes are marked in italics and non-conserved changes in lower case. Conserved

changes are considered as substitutions within the amino acid homology groups FWY, MILV, RKH, ED, NQ, ST and PAG. Gaps introduced for optimal alignment are represented by asterisks.

5 The amino acid sequence of the Ac-AMPs shows striking similarity to the cysteine/glycine-rich domains of chitin-binding proteins, such as chitinases, chitin-binding lectins, and hevein. However, the Ac-AMPs also contain unique features.

10 Sequence alignment of Ac-AMP2 and the N-terminus of a basic chitinase from tobacco (Figure 4B) showed 14 identical amino acids and 5 conserved changes in the first 30 residues. A single gap of four amino acids had to be introduced in the N-terminal

15 portion of Ac-AMP2 to allow optimal alignment with the chitin-binding proteins. After introduction of this gap, all of the cysteine residues appeared at invariant positions.

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EXAMPLE 6

Chitin-binding activity of the Ac-AMPs.

Because of the similarity at the amino acid sequence level between the Ac-AMPs and

25 chitin-binding proteins, the ability of the amaranth antifungal proteins to bind on a chitin substrate was investigated.

Micro-columns packed with chitin were loaded with either Ac-AMP1 or Ac-AMP2 and subsequently

30 eluted at neutral pH and low pH (pH 2.8). Chitin was prepared by N-acetylation of chitosan (Sigma, St Louis, MO) by the method of Molano et al (1977, Anal Biochem, 83:648-656). Protein samples (50 µg) dissolved in 1 ml phosphate buffered saline (pH 7)

were applied on the chitin micro-column (2.5 x 6 mm) and recycled three times over the column. The column was eluted five times with 1 ml phosphate buffered saline (PBS) and once with 1 ml 100 mM acetic acid (pH 2.8). Fractions (1 ml) of the eluate were desalted and concentrated by reversed-phase chromatography and finally redissolved in 50 μ l sample buffer for SDS-PAGE analysis.

Figure 5 shows results of the SDS-PAGE analysis after this affinity chromatography of Ac-AMP1 (lanes 1-4) and Ac-AMP2 (lanes 5-8). Lanes 1 and 5 are the antifungal proteins at equivalent amounts as those loaded on the columns; lanes 2 and 6 are the fractions passed through the column; lanes 3 and 7 are the fractions eluted with PBS (pH 7); lanes 4 and 8 are the fractions eluted with 100 mM acetic acid (pH 2.8). It can be seen that the Ac-AMPs were absent from the fraction passed through the column and from the neutral pH washings, but instead were recovered in the low pH desorption buffer. These results indicate that both Ac-AMP1 and Ac-AMP2 exhibit affinity toward chitin.

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EXAMPLE 7

Stability of the antifungal activity of the Ac-AMPs.

Tests for antifungal activity were performed with 20 μ l samples diluted five-fold with growth medium containing Fusarium culmorum spores, according to the assay method given in Example 2. Untreated control samples consisted of the test proteins at 500 μ g/ml in 10 mM sodium phosphate

buffer (pH 7). For digestions, different proteases were added at 100 μ g/ml and incubated at 37°C for 16 hours. Heat stability tests were performed by heating aliquots of the test proteins for 10 minutes at different temperatures up to 100°C. pH stability was tested by incubation of test proteins for 1 hour in either 20 mM glycine-HCl (pH 2) or glycine-NaOH (pH 11) and subsequent dialysis for 16 hours against 10 mM sodium phosphate buffer (pH 7) using benzoylated cellulose tubing. Reduction was done by addition of dithiothreitol at 30 mM and Tris-HCl (pH 8.6) at 300 mM. The reagents were removed by reversed-phase chromatography. Any of the control treatments containing only the reagents proved negative for antifungal activity after the dialysis or reversed-phase chromatography steps.

The antifungal activity of the Ac-AMPs was resistant to digestion by proteinase K, pronase E, chymotrypsin or trypsin. Moreover, the Ac-AMPs were not affected by heat treatments at up to 100°C for 10 minutes nor by exposure to pH conditions as extreme as pH 2 or pH 11. Reduction of their cysteine residues by dithiothreitol, however, completely abolished the antifungal activity.

The proteins are remarkably stable, since their biological activity is unaffected by protease treatments or by exposure to extreme temperatures and pH conditions. This stability may be due to a compact globular structure maintained by the relatively high number of disulphide linkages. These disulphide linkages are essential for biological activity.

EXAMPLE 8

Antifungal potency of the Ac-AMPs.

The antifungal potency of the Ac-AMPs was assessed on fourteen different plant pathogenic fungi, using the assay described in Example 2. Growth of fungi, collection and harvest of fungal spores, and preparation of mycelial fragments were done as previously described (Broekaert et al, 1990, FEMS Microbiol Lett, 69:55-60). The following fungal strains were used: Alternaria brassicola MUCL 20297, Ascochyta pisi MUCL 30164, Botrytis cinerea MUCL 30158, Cercospora beticola strain K897, Colletotrichum lindemuthianum MUCL 9577, Fusarium culmorum IMI 180420, Mycosphaerella fijiensis var fijiensis IMI 105378, Phytophthora infestans, Rhizoctonia solani CBS 207-84, Sclerotinia sclerotium MUCL 30163, Septoria nodorum MUCL 30111, Trichoderma hamatum MUCL 29736, Verticillium dahliae MUCL 19210, and Venturia inaequalis MUCL 15927.

For C beticola, R solani, S sclerotium, S nodorum, M fijiensis and P infestans, mycelial fragments were used as inoculum. All other fungi were inoculated as spores.

Figure 6 shows the dose-response curves of fungal growth inhibition measured at varying concentrations of Ac-AMP1 (panel A) and Ac-AMP2 (panel B) using the following test fungi: A brassicola (*); A pisi (x); B cinerea (+); C lindemuthianum (□); F culmorum (■); V dahliae (▲).

The antifungal activity of the Ac-AMPs on the fourteen plant pathogenic fungi listed above was compared to that of two known chitin-binding proteins, nettle lectin and pea chitinase. Table 1

summarises the results: IC_{50} is the concentration ($\mu g/ml$) required for 50% growth inhibition after 48 hours of incubation. The IC_{50} values for the slow growing fungi S nodorum and V inaequalis were measured after 5 and 15 days of incubation respectively. The nettle lectin (or Urtica dioica agglutinin, UDA) was isolated from stinging nettle (Urtica dioica) rhizomes as previously described (Peumans et al, 1983, FEBS Lett, 177:99-103). Chitinase was isolated from pea pods by the method of Mauch et al (1988, Plant Physiol, 87:325-333).

TABLE 1

Antifungal activity of
Ac-AMPs, nettle lectin and pea chitinase

Fungus	IC_{50} ($\mu g/ml$)			
	Ac-AMP1	Ac-AMP2	UDA	chitinase
<u>A brassicola</u>	7	4	200	400
<u>A pisi</u>	8	8	1000	>500
<u>B cinerea</u>	10	8	>1000	>500
<u>C beticola</u>	0.8	0.8	ND	ND
<u>C lindemuthianum</u>	8	8	20	>500
<u>F culmorum</u>	2	2	>1000	>500
<u>M fijiensis</u>	3	ND	4	ND
<u>P infestans</u>	12	ND	4	ND
<u>R solani</u>	30	20	30	ND
<u>S sclerotianum</u>	20	10	ND	ND
<u>S nodorum</u>	20	20	ND	ND
<u>T hamatum</u>	7	3	90	1.5
<u>V dahliae</u>	6	5	80	500
<u>V inaequalis</u>	ND	3	1000	ND

ND = not determined

The concentration of Ac-AMP protein required for 50% growth inhibition after 48 hours of incubation (IC_{50}) varied from 0.8 to 30 $\mu\text{g/ml}$, depending on the test organism. The antifungal potency of Ac-AMP1 was almost identical to that of Ac-AMP2.

The Ac-AMPs are potent inhibitors of all fourteen fungi tested in this study. Their specificity is comparable to that of wheat thionin which also typically inhibits fungal growth with IC_{50} values between 1 and 10 $\mu\text{g/ml}$ (Cammue et al, 1992, J Biol Chem, 267, 2228-2233). Relative to other chitin-binding proteins, such as the nettle lectin or chitinase, the Ac-AMPs have much higher specific activities. The nettle lectin only inhibits 6 out of 11 fungi at concentrations below 100 $\mu\text{g/ml}$, whereas at this concentration the pea chitinase is only inhibitory to 1 out of 7 tested fungi.

The unique properties of the Ac-AMPs as potent inhibitors of fungal growth in vitro suggest that they may play a role in the defence of seeds or seedlings against invasion by fungal organisms.

25

EXAMPLE 9

Effect of ions on antifungal activity.

The specific activity of the Ac-AMPs was found to be strongly dependent on the ionic constitution of the growth medium. Figure 7 shows the dose-response curves of Ac-AMP1 (panel A) and Ac-AMP2 (panel B) on B cinerea in a low ionic strength synthetic growth medium, with and without different additions of KCl or CaCl_2 . The antagonistic effect of K^+ and Ca^{2+} on growth

inhibition of B cinerea caused by the Ac-AMPs is obvious. In the reference medium (■), containing 2.5 mM monovalent cations and 0.1 mM divalent cations, Ac-AMP1 and Ac-AMP2 had IC_{50} values of 2.2 and 1.6 $\mu\text{g/ml}$ respectively. Administering KCl at 10 mM (x) to this medium did not significantly affect the dose-response curves, whereas KCl at concentrations of 50 mM (□) increased the IC_{50} values by about three-fold. CaCl_2 had a much more dramatic antagonistic effect. When supplemented at 1 mM (*) to the reference medium, CaCl_2 caused a five to six-fold increase of the IC_{50} values. At 5 mM CaCl_2 (+) the drops in specific activity were more than 50-fold. The antagonistic effect of other salts with monovalent cations, such as NaCl and NH_4Cl , was similar to that of KCl, whereas the effect of the salts with divalent cations, MgCl_2 and BaCl_2 , was similar to that of CaCl_2 .

These results show that the antifungal activity of the Ac-AMPs is strongly reduced by the presence of inorganic salts. The antagonistic effect of salts is primarily due to the cations; divalent cations are more potent antagonists than monovalent cations.

EXAMPLE 10

Effect of Ac-AMPs on bacteria.

Antibacterial activity was measured as described in Example 2. The following bacterial strains were used: Bacillus megaterium ATCC 13632, Erwinia carotovora strain 3912, Escherichia coli strain HB101 and Sarcina lutea ATCC 9342. The antibacterial effect of the Ac-AMPs was assessed by adding serial dilutions of the proteins to

bacterial suspensions. The highest test concentration was 500 $\mu\text{g/ml}$ (final concentration). Results are shown in Table 2.

5

TABLE 2

Antibacterial activity of the Ac-AMPs

Bacteria	IC ₅₀ ($\mu\text{g/ml}$)	
	Ac-AMP1	Ac-AMP2
<u>B megaterium</u>	40	10
<u>S lutea</u>	250	40
<u>E carotovora</u>	no inhibition	
<u>E coli</u>	no inhibition	

15

The Ac-AMPs inhibited growth of the gram-positive bacteria, B megaterium and S lutea. However, the Ac-AMPs (at 500 $\mu\text{g/ml}$) did not inhibit growth of the gram-negative bacteria E carotovora and E coli.

20

EXAMPLE 11

Effect of the purified antimicrobial proteins on cultured human cells.

The Ac-AMPs were evaluated for their potential toxic effects on mammalian cells.

25

Human cell toxicity assays were performed either on umbilical vein endothelial cells (Alessi et al, 1988, Eur J Biochem, 175, 531-540) or skin-muscle fibroblasts (Van Damme et al, 1987, Eur J Immunol, 17, 1-7) cultured in 96-well microplates. The growth medium was replaced by 80 μl of serum-free medium (Optimem 1 for endothelial cells or Eagle's minimal essential medium (EMEM) for fibroblasts, both from GIBCO), to

30

which 20 μ l of a filter-sterilised test solution was added. The cells were further incubated for 24 hours at 37°C under a 5% CO₂ atmosphere with 100 % relative humidity. The viability of the cells was assessed microscopically after staining with trypane blue (400 mg/l in phosphate buffered saline, PBS) for 10 minutes. Alternatively, cells were stained with neutral red (56 mg/l in PBS) for 2 h at 37°C. Cells were lysed in acidic ethanol (100 mM sodium citrate, pH 4, containing 50% ethanol) and scored for release of the dye by microspectrophotometry at 540 nm.

When added at up to 500 μ g/ml to either cultured human umbilical vein endothelial cells or human skin-muscle fibroblasts, neither Ac-AMP1 nor Ac-AMP2 affected cell viability after 24 h of incubation. In contrast, β -purothionin administered at 50 μ g/ml decreased the viability of both cell types by more than 90%.

20

EXAMPLE 12

Molecular cloning of Ac-AMP2 cDNA.

Fully matured seeds of Amaranthus caudatus were collected from outdoor grown plants, immediately frozen in liquid nitrogen and stored at -80°C. Total RNA was extracted from 5 g of pulverised seeds by the method of De Vries et al (1988, Plant Molecular Biology Manual, B6, 1-13) using 6 ml of a 1:2 phenol:RNA extraction buffer mixture and 2 ml of chloroform per g tissue. Poly (A)⁺ RNA was purified by oligo (dT)-cellulose affinity chromatography as described by Silflow et al (1979, Biochemistry, 18, 2725-2731) yielding about 7 μ g of poly (A)⁺ RNA. Double-stranded cDNAs

were prepared from 1.5 μ g of poly (A)⁺ RNA according to Gubler and Hoffman (1983, Gene, 25, 263-269) and ligated to EcoRI/NotI adaptors using the cDNA Synthesis Kit of Pharmacia. The cDNAs
5 were cloned into the λ ZAP II phage vector (Stratagene) and packaged in vitro with the Gigapack II Gold packaging system (Stratagene) according to the manufacturer's instructions.

A DNA probe for screening of the cDNA library
10 was produced by polymerase chain reaction (PCR) as follows. Two degenerate oligonucleotides were synthesised: OWB13 (5'GTNGGNGARTGKGTMGNGG) and OWB14 (5'CCRCARTAYTTNGGNCCYTTNCC). OWB13 corresponds to amino acids 1 to 7 of Ac-AMP1 and
15 has a sense orientation. OWB14 corresponds to amino acids 22 to 29 of Ac-AMP1 and has an antisense orientation. PCR was performed with the Taq polymerase under standard conditions (Sambrook et al, 1989, Molecular Cloning, Cold Spring Harbour
20 Lab Press) using OWB13 and OWB14 as amplimers and 25ng of cDNA as target DNA. The temperature programme included an initial step at 94°C for 5 min, 30 cycles (94°C for 1 min; 45°C for 2 min; 72°C for 3 min) and a final step at 72°C for 10
25 min. The 100 bp PCR amplification product was purified on a 3% agarose (NuSieve, FMC) gel and reamplified by PCR under the same conditions except that the reaction mixtures contained 130 μ M dTTP and 70 μ M digoxigenin-11-dUTP instead of 200 μ M
30 dTTP. The digoxigenin-labelled PCR product was purified on a 3% NuSieve agarose gel.

About 100,000 plaque forming units of the λ ZAP II cDNA library were screened with the digoxigenin-labelled PCR product by in situ plaque

hybridisation using nylon membranes (Hybond -N, Amersham). Membranes were air-dried and DNA was crosslinked on the membranes under UV light (0.15 J/cm^2). Hybridisation was performed for 16 hours at 65°C in 5 x SSC, 1% blocking reagent (Boehringer Mannheim), 0.1% N-lauroylsarcosine, 0.02% sodium dodecylsulphate containing 10 ng/ml of heat denatured digoxigenin-labelled probe.

Non-specifically bound probe was removed by rinsing twice for 5 min in 2 x SSC/0.1% SDS at 25°C and twice for 15 min in 0.1 x SSC/0.1% SDS at 60°C. Detection of the probe was done using anti-digoxigenin antibodies linked to alkaline phosphatase (Boehringer Mannheim) and its substrate 5-bromo-4-chloro-3-indolyl phosphate (Boehringer Mannheim) according to the manufacturer's instructions. Positive plaques were purified by two additional screening rounds with the same probe under the same conditions. Inserts from purified plaques were excised in vivo into the pBluescript phagemid form with the aid of the helper phage R408, according to the instructions of Stratagene. Nucleotide sequencing was done with an ALF automated sequencer (Pharmacia) using fluoresceine-labelled M13 forward and reverse primers (Pharmacia). Sequence analysis was performed by the Intelligenetics PC-gene software.

Inserts from ten different positive clones were released by EcoRI digestion and their sizes compared by agarose electrophoresis. The clone with the longest insert (AC1) was subjected to nucleotide sequence analysis. AC1 is 590 nucleotides long and contains an open reading frame of 86 amino acids. The 25 amino-terminal amino

acids have a predictable signal peptide structure obeying the (-1,-3)-rule (Von Heijne, 1985, Mol Biol, 184, 99-105). The deduced amino acid sequence of the region following the putative
5 signal peptide is identical to the 30-amino acid sequence of mature Ac-AMP2 as determined by protein sequencing. In addition, the mature protein domain is extended by a 31-amino acids carboxy-terminal domain. This carboxy-terminal extension may play a
10 role in the subcellular targeting of Ac-AMP2. AC1 has 45-nucleotide and 284-nucleotide untranslated regions at the 5' and 3' end, respectively. The 3' end untranslated region is not terminated by a poly(A) tail, indicating that AC1 is not a full
15 length cDNA clone.

All of the other nine sequenced positive clones had deduced amino acid sequences identical to that of AC1. They differed from each other by various degrees of truncation at the 5' or 3' end.
20 The fact that the 10 sequenced clones all contained an arginine at position 30 of the mature protein domain suggests that Ac-AMP1 and Ac-AMP2 are both derived from the same precursor preprotein.

The carboxy-terminal extension peptide did not
25 show relevant homology either at the amino acid or nucleotide level with any of the entries from the Swiss-Prot (release 20) or EMBL gene bank (release 29), respectively. The structure of the Ac-AMP2 cDNA thus appears to be unique and obviously
30 different from that of known genes encoding chitin-binding proteins.

Figure 8 shows the nucleotide sequence and deduced amino acid sequence of clone AC1. The putative signal sequence is underlined and the

sequence of mature Ac-AMP2 is boxed. The stop codon is marked with an asterisk.

EXAMPLE 13

5 Construction of the expression vectors pAC11 and pAC12.

Two different expression cassettes were constructed based on two portions of the insert of clone AC1.

10 The first expression vector (pAC11, as shown in Figure 9A) contains the full coding region of Ac-AMP2 cDNA flanked at its 5' end by the strong constitutive promoter of the 35S RNA of cauliflower mosaic virus, CaMV35S (Odell et al, 1985, Nature, 313, 810-812) with a duplicated enhancer element to
15 allow for high transcriptional activity (Kay et al, 1987, Science, 236, 1299-1302). The coding region of Ac-AMP2 cDNA is flanked at its 3' side by the CaMV35S polyadenylation sequence. The plasmid
20 backbone of this vector is the phagemid pUC120 (Vieira and Messing, 1987, Methods Enzymol, 153, 3-11).

pAC11 was constructed from clone AC1 as follows. Clone AC1 consists of the Ac-AMP2 cDNA
25 (shown in Figure 8) cloned into the EcoRI site of pBluescript SK(+) (from Stratagene) such that the 5' end faces the M13 universal primer binding site. AC1 was digested with EcoRV (cuts within the SK polylinker of pBluescript) and NheI (cuts
30 internally in the Ac-AMP2 cDNA sequence at base position 315 which is 9 bases downstream of the stop codon). The EcoRV/NheI 332 bp fragment was subcloned into the expression vector pFAJ3002 which was pre-digested with SmaI and XbaI. pFAJ3002 is a

derivative of the expression vector pFF19 (Timmermans et al, 1990, J Biotechnology, 14, 333-344) in which the unique EcoRI site is replaced by a HindIII site.

5 The second expression vector (pAC12, shown in Figure 9B) contains an open reading frame coding for the signal peptide and mature domain of the Ac-AMP preprotein. This open reading frame is flanked at its 5' side by the duplicated CamV35S
10 promoter and at its 3' side by the CamV35S polyadenylation sequence.

pAC12 was constructed as follows. A 216 bp fragment was amplified by polymerase chain reaction using AC1 as DNA template and OWB32, OWB33 as sense
15 and antisense primers respectively. The primer OWB32 (5'AATTGGATCCAGTCAAGAGTATTAATTAGG) corresponds to nucleotides 17 to 36 of Ac-AMP2 cDNA and introduces a BamHI site at the 5' end of the PCR amplification product. The primer OWB33
20 (5'AATTGTCGACTCAACGGCCACAGTACTTTGGGCC) corresponds to nucleotides 190 to 210 of Ac-AMP2 cDNA and links an inframe stop codon and a SalI site to the 3' end of the PCR products. Both OWB32 and OWB33 have a 4 bp random sequence at the 5' end prior to the
25 restriction site. The PCR product was digested with BamHI and SalI and subsequently subcloned into the expression vector pFAJ3002, previously digested with the same restriction enzymes.

30

EXAMPLE 14

Construction of the plant transformation vectors pAC111 and pAC112.

The expression vector pAC11 and pAC12 were digested with HindIII and the fragments containing

the Ac-AMP2 cDNA expression cassettes were subcloned into the unique HindIII site of pBin19Ri. pBin19Ri is a modified version of the plant transformation vector pBin19 (Bevan, 1984, Nucleic Acids Research, 12:22, 8711-8721) wherein the unique EcoRI and HindIII sites are switched and the defective npt II expression cassette (Yenofsky et al, 1990, Proc Natl Acad Sci USA, 87:3435-3439) is replaced by the npt II expression cassette described by An et al (1985, EMBO J, 4:277-284).

The plant transformation vector containing the pAC11 expression cassette was designated pAC111, while the transformation vector containing the pAC12 expression cassette was designated pAC112. The structure of the two transformation vectors is shown in Figure 10.

EXAMPLE 15

Plant Transformation.

The disarmed Agrobacterium tumefaciens strain LBA4404 [pAL4404] (Hoekema et al, 1983, Nature, 303:179-180) may be transformed by either of the vectors pAC111 or pAC112, using the method of de Framond A et al (Biotechnology, 1:262-9).

Tobacco transformation may be carried out using leaf discs of Nicotiana tabacum Samsun based on the method of Hozsch RB et al (1985, Science, 227, 1229-31) and co-culturing with Agrobacterium strains containing pAC111 or pAC112.

Co-cultivation may be carried out under selection pressure of 100 μ g/ml kanamycin. Transgenic plants (transformed with pAC111 or pAC112) may be regenerated on media containing 100 μ g/ml kanamycin. These transgenic plants may be analysed

for expression of the newly introduced genes using standard Western blotting techniques. Transgenic plants may also be analysed for increased resistance to fungal or bacterial diseases.

5 Plants capable of constitutive expression of the introduced genes may be selected and self-pollinated to give seed. The progeny of the seed exhibiting stable integration of the Ac-AMP genes would be expected to show typical Mendelian
10 inheritance patterns for the Ac-AMP genes.

CLAIMS

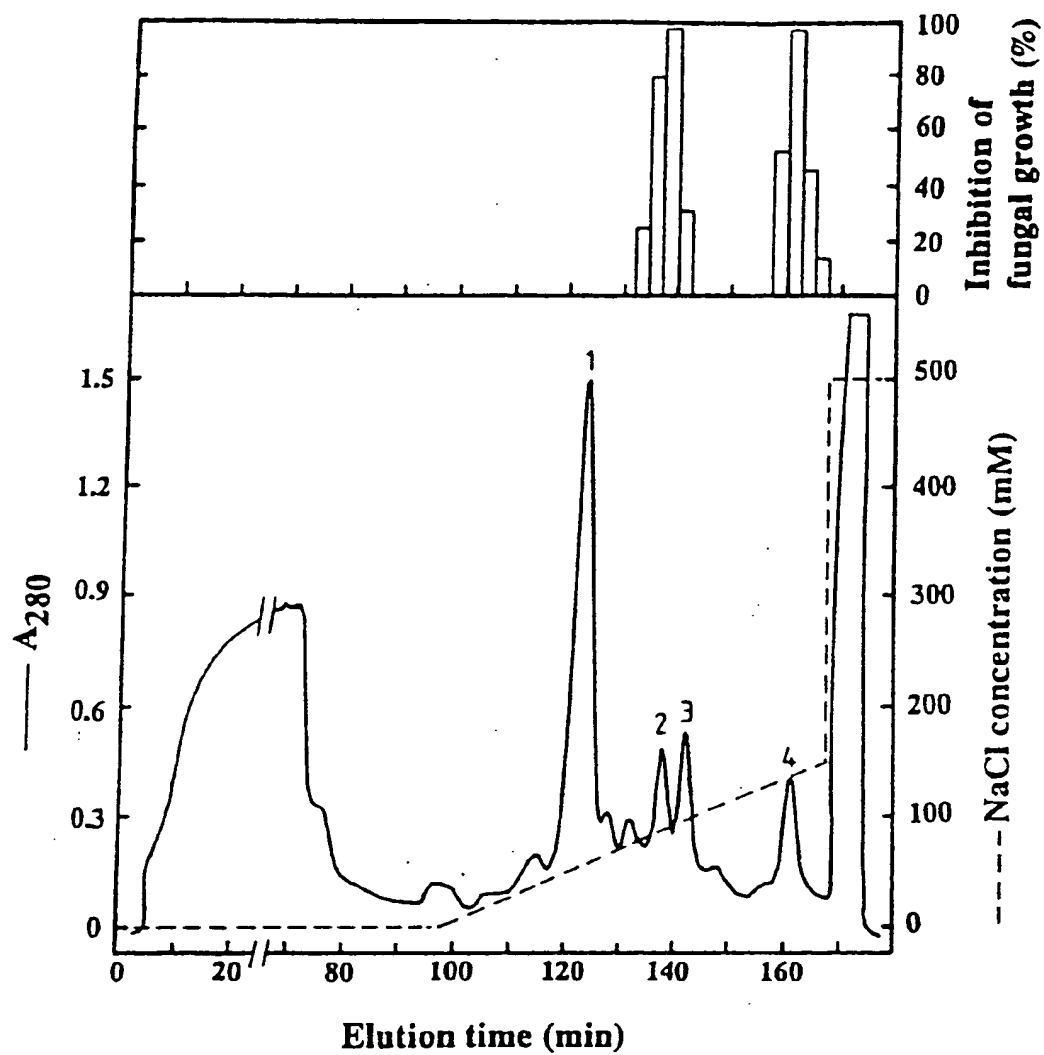
- 1 Antimicrobial proteins isolated from
5 Amaranthus seed.
- 2 The pure protein Ac-AMP1, capable of being
isolated from Amaranthus seed.
- 10 3 The pure protein Ac-AMP2, capable of being
isolated from Amaranthus seed.
- 4 Pure proteins having the amino acid sequence
of proteins as claimed in any of claims 1-3.
15
- 5 Protein as claimed in claim 4 which is
synthetic.
- 6 A recombinant DNA sequence coding for a
20 protein as claimed in any of claims 1-5.
- 7 A vector containing a DNA sequence as claimed
in claim 6.
- 25 8 A vector as claimed in claim 7 containing a
DNA sequence isolated from a plant genome.
- 9 A biological system including DNA as claimed
in claim 6 which allows expression of the
30 encoded protein.
- 10 A biological system as claimed in claim 9
which is a micro-organism.

- 11 A biological system as claimed in claim 9
which is a plant.
- 5 12 Plants transformed with recombinant DNA as
claimed in claim 6.
- 13 Plants transformed with a recombinant DNA
sequence coding for the protein Ac-AMP1.
- 10 14 Plants transformed with a recombinant DNA
sequence coding for the protein Ac-AMP2.
- 15 15 Protein derived from expression of the DNA as
claimed in claim 6.
- 16 Antimicrobial protein produced by expression
of recombinant DNA within plants as claimed in
any of claims 12-14.
- 20 17 An antimicrobial composition containing one or
more of the proteins as claimed in any of
claims 1-5 or claims 15-16.
- 25 18 A process of combating fungi or bacteria which
comprises exposing them to the proteins or
compositions as claimed in any of claims 1-5
or claims 15-17.
- 30 19 An extraction process for producing biocidally
active proteins as claimed in any of claims
1-5 or claims 15-16 from organic material
containing them which comprises submitting the
organic material to maceration and solvent
extraction.

- 20 An extraction process as claimed in claim 19
where the protein is subsequently purified by
centrifugation, chromatography and dialysis.
- 5 21 An extraction process as claimed in either
claim 19 or claim 20 where the organic matter
comprises seeds of Amaranthus.
- 10 22 An extraction process as claimed in either
claim 19 or claim 20 where the organic matter
comprises a micro-organism as claimed in claim
10.

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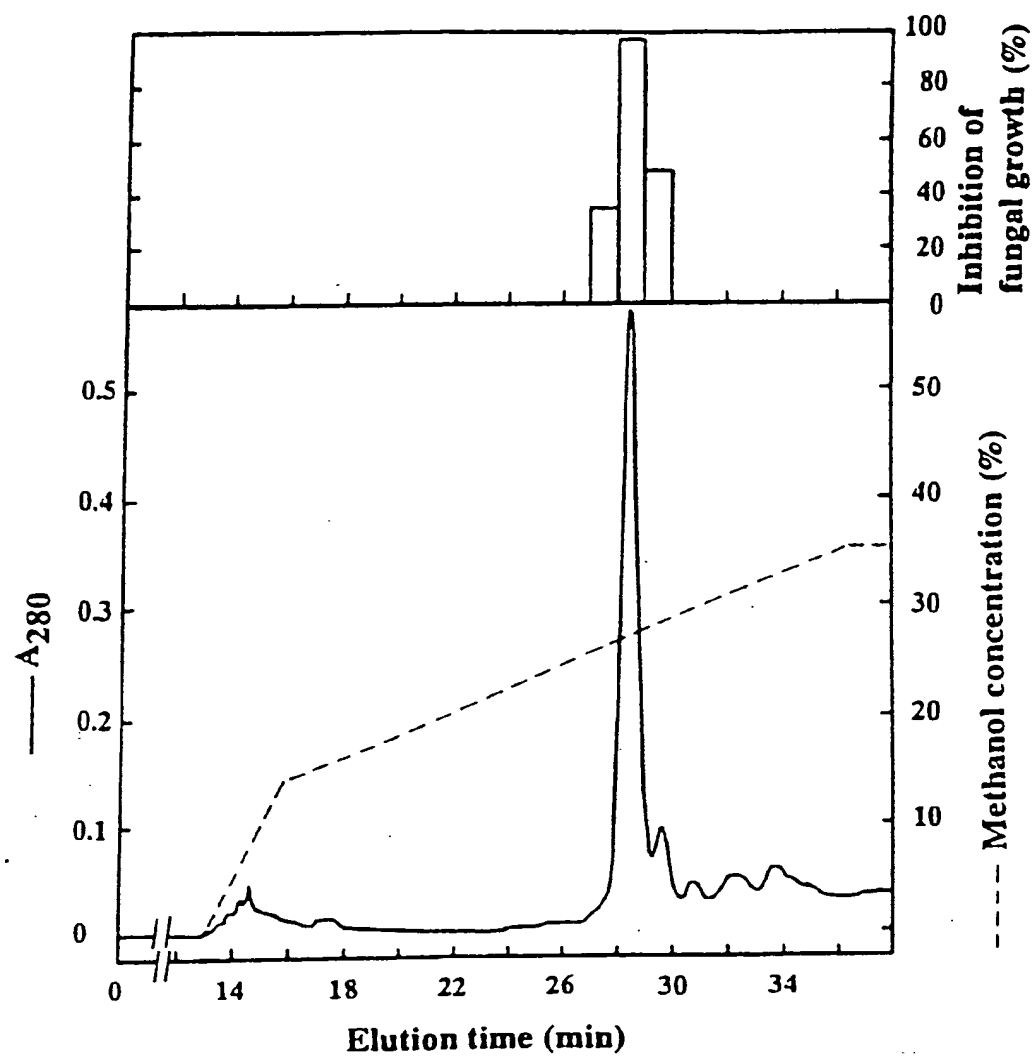
FIG. 1



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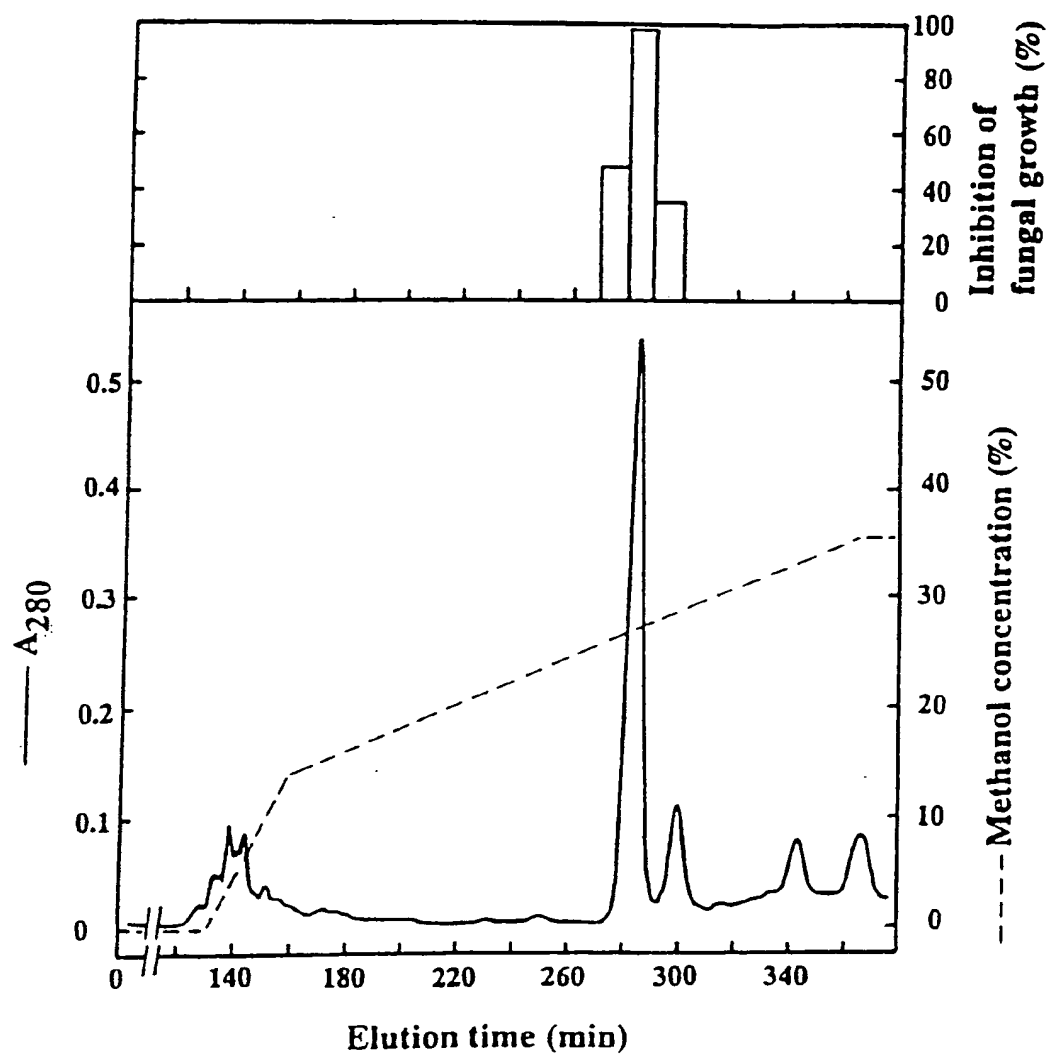
FIG. 2A



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FIG. 2B



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FIG. 3

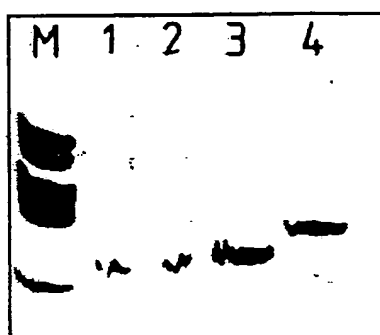
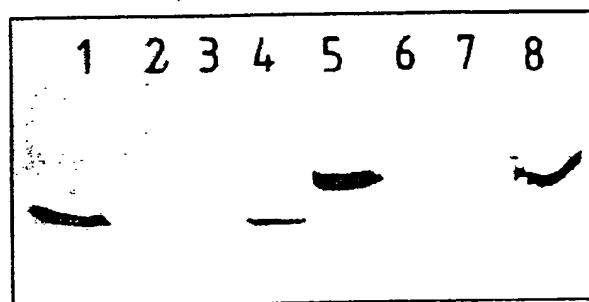


FIG. 5



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FIG. 4

A

Ac-AFP1

V G E C V R G R C P S G M C C S Q F G Y C G K G P K Y C G

Ac-AFP2

V G E C V R G R C P S G M C C S Q F G Y C G K G P K Y C G R

B

Bean chitinase

* E Q C G S Q A G G A R C A S G L C C S K F G W C G N T N D Y C G P G * N C Q S * Q C P G G

Tobacco chitinase

* E Q C G R Q A G G A I C P G G n C C S q F G W C G s T t D Y C G P G * * C Q S * Q C * G G

Hevein

* E Q C G R Q A G G k I C P n n L C C S q W G W C G s T d E Y C s P d h N C Q S * W C k d s

Wheat lectin

* q r C G e Q G s n n e C P n n L C C S q Y G Y C G m g g D Y C G K G * * C Q d g a C w t s

Nettle lectin

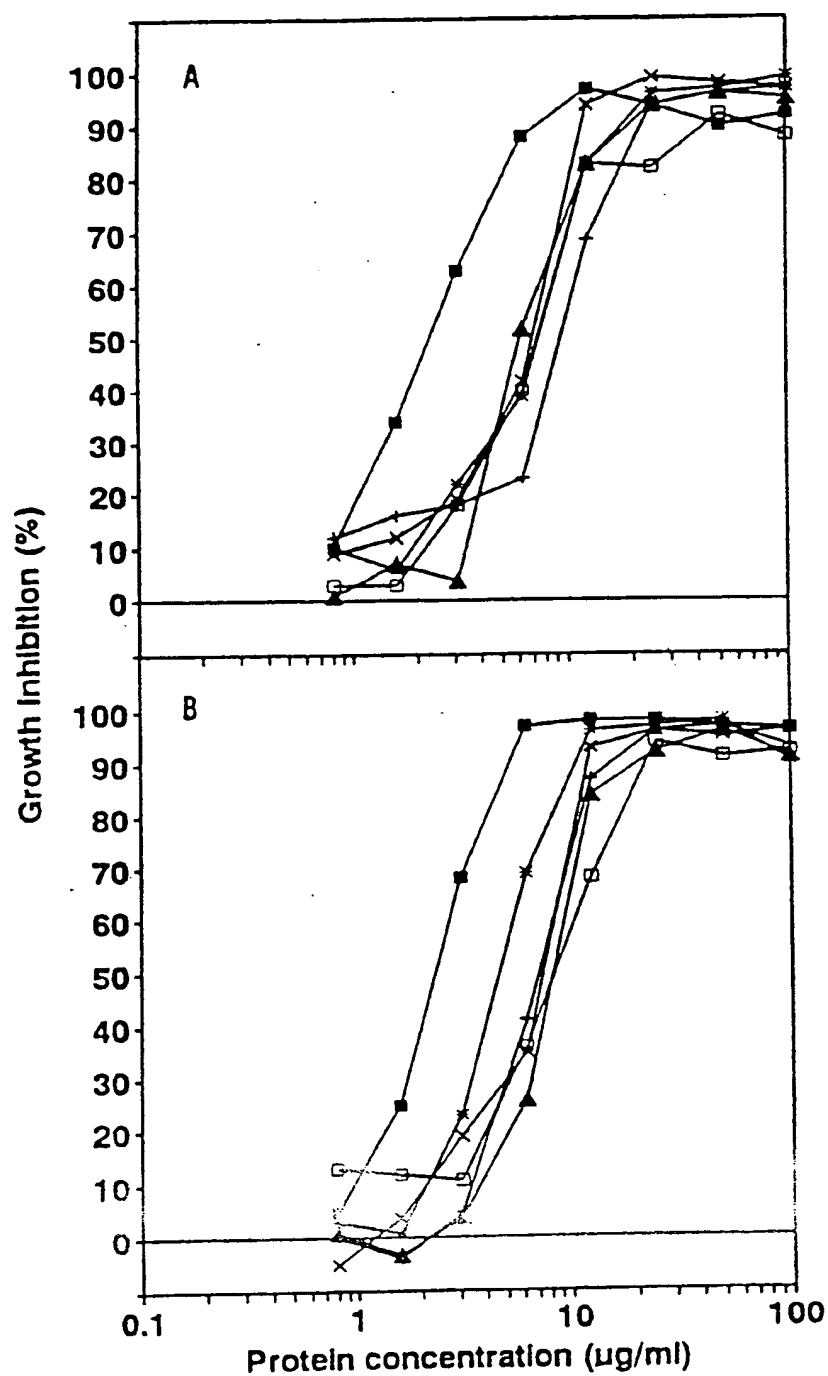
* q r C G S Q G G G G t C P a I r C C S i W G W C G a S s p Y C

Ac-AFP2

v g e C v r * * * G R C P S G M C C S q F G Y C G K g p k Y C G r

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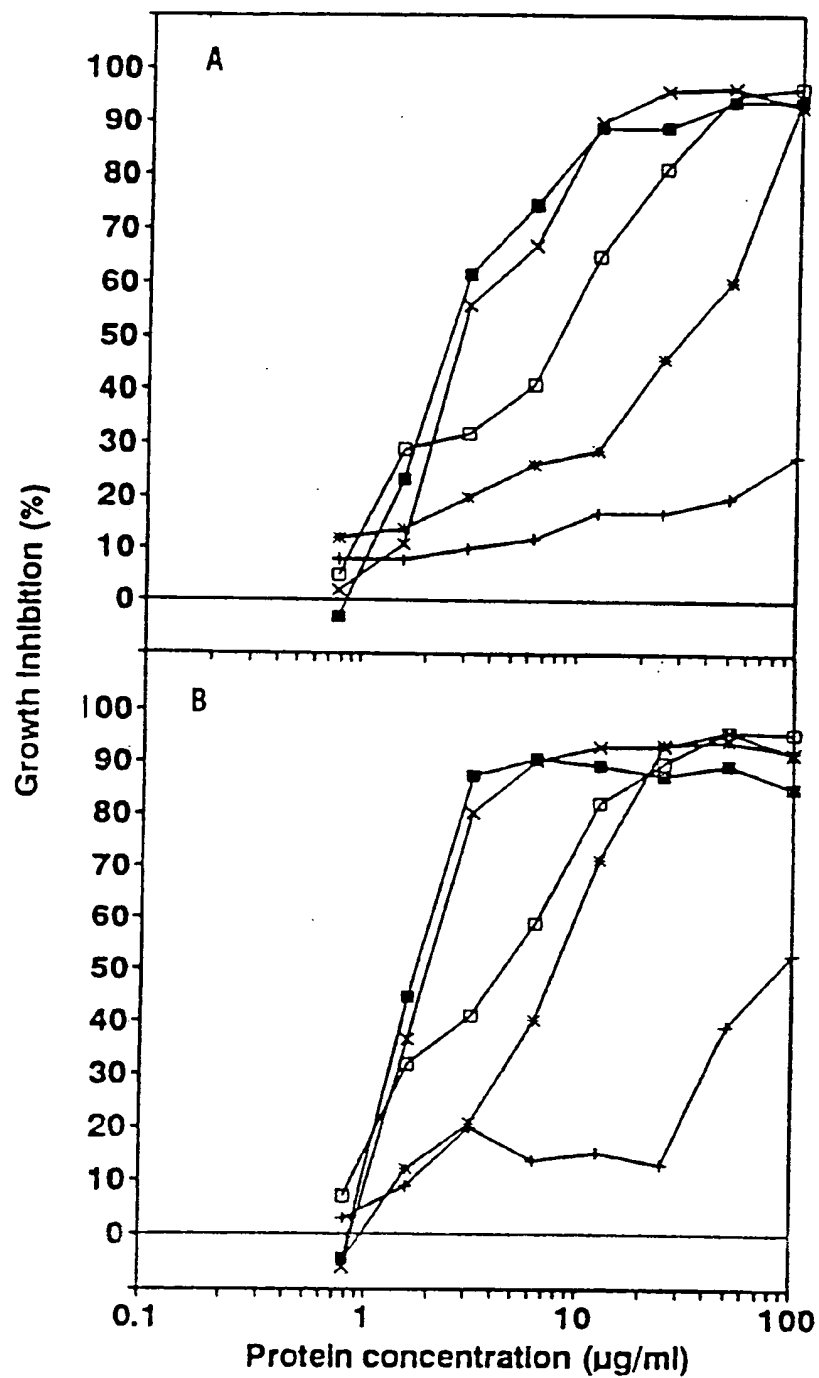
FIG. 6



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FIG. 7



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FIG.8

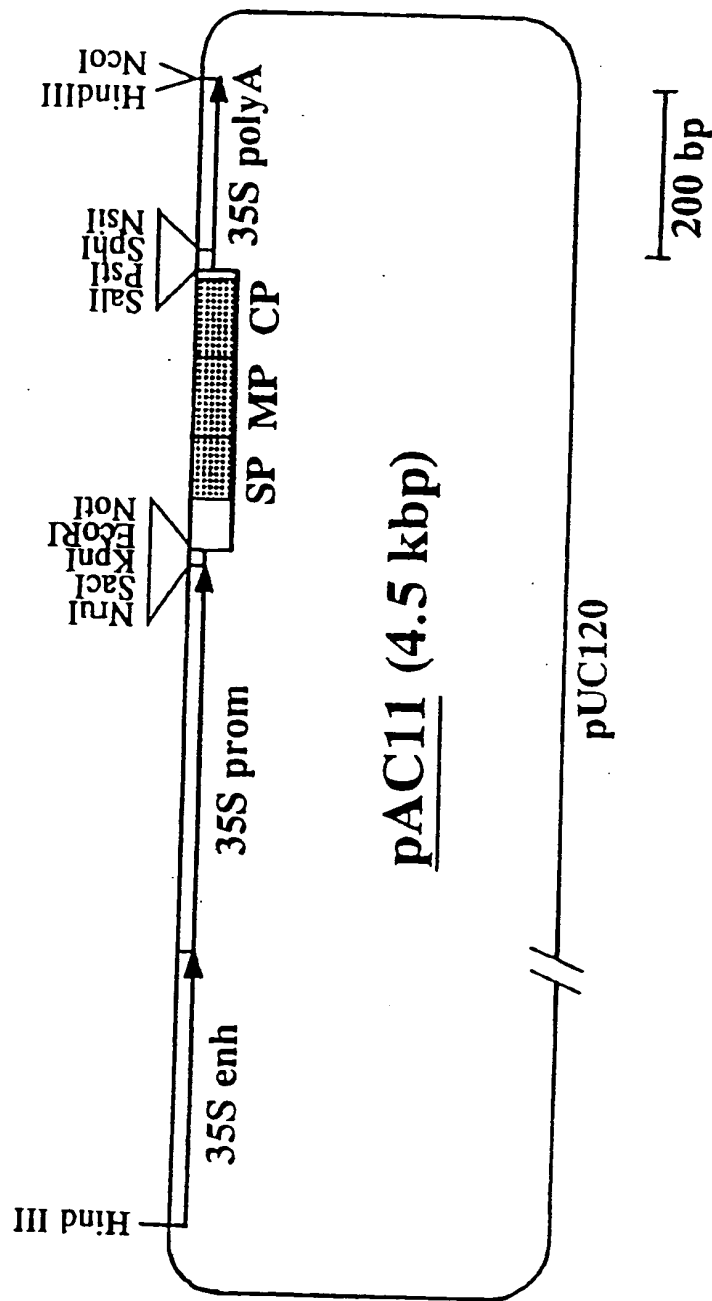
Nucleotide sequence and deduced amino acid sequence of Ac-AMP cDNA

1	CAAAAAAAAAATAAGTCAAGAGTATTAAATTAGGTGAGAAAAAATGGTGAACATGAAG	<u>M V N M K</u>
61	TGTGTTGCATTGATAGTTATAGTTATGATGGCGTTTATGATGGTGGATCCATCAATGGGA	<u>C V A L I V I V M M A F M M V D P S M G</u>
121	GTGGGAGAATGTGTGAGAGGACGTTGCCCAAGTGGGATGIGTIGCAGTCAGTTTGGGTAC	<u>V G E C V R G R C P S S G M C C S Q F G Y</u>
181	TGTGGTAAAGGCCCAAGTACTGTGGCCGTCGCCAGTACTACTGTGGATCACCACGCTGAT	<u>C G K G P K Y C G R</u> A S T T V D H Q A D
241	GTTGCTGCCACCAAACTGCCAAGAATCCTACCGATGCTAAACTTGCIGGTGCTGGTAGT	V A A T K T A K N P T D A K L A G A G S
301	CCATGAAAGTAGTAGCTAGGTTACGTTGGATTACCAAGCCGTCGCCAGTACTACTG	P *
361	TGGCCGTGCCAGTACTAATGTTCTCTTATATGTCTGAAATAAGCTCCTATATAAATACTA	
421	GIATCTTGATGTAATGGAGTATTTTCATTTTGTGTTTATTTAGTTATGATCGTGACTTC	
481	CTTGTTGGTTTAACTTGTATATTTGTAATGCATCTTAAATGCTGTCTCAATAATTGA	
541	TGIATTAACACTTGTGTTTGTGTTTAAATACATACTAAGTGTGTAAATTC	

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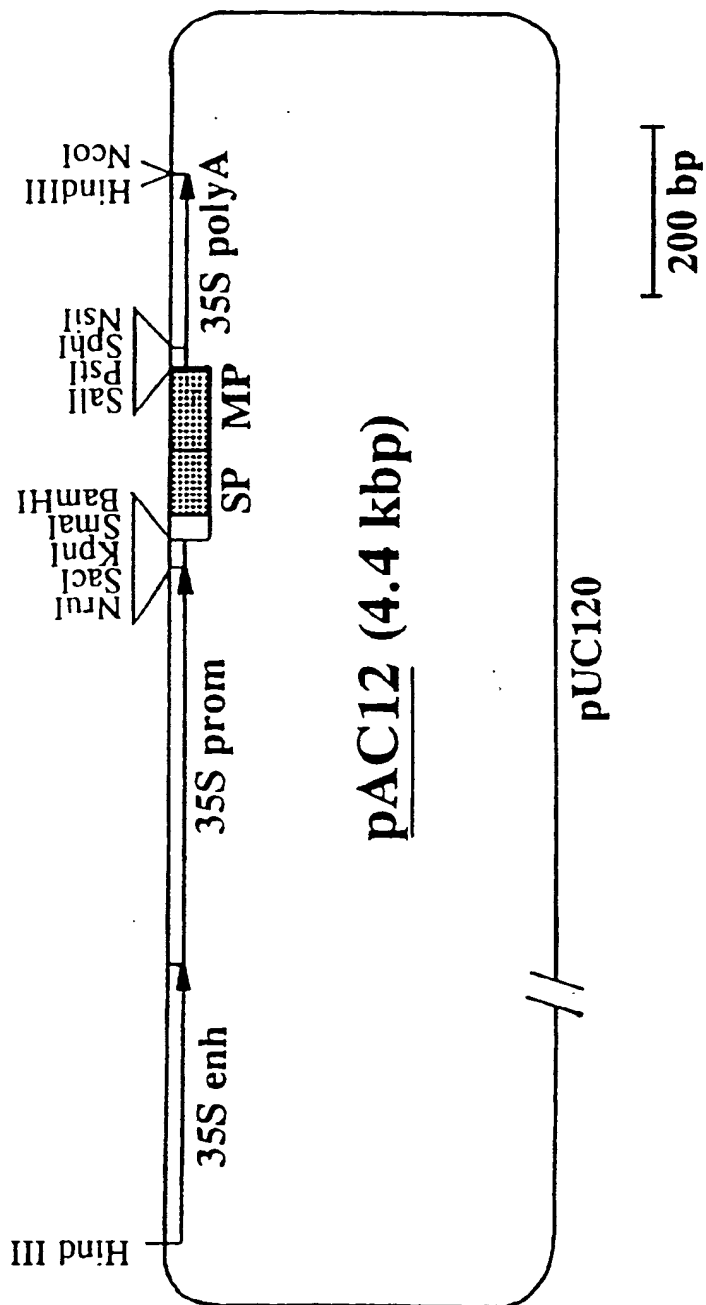
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FIG. 9A



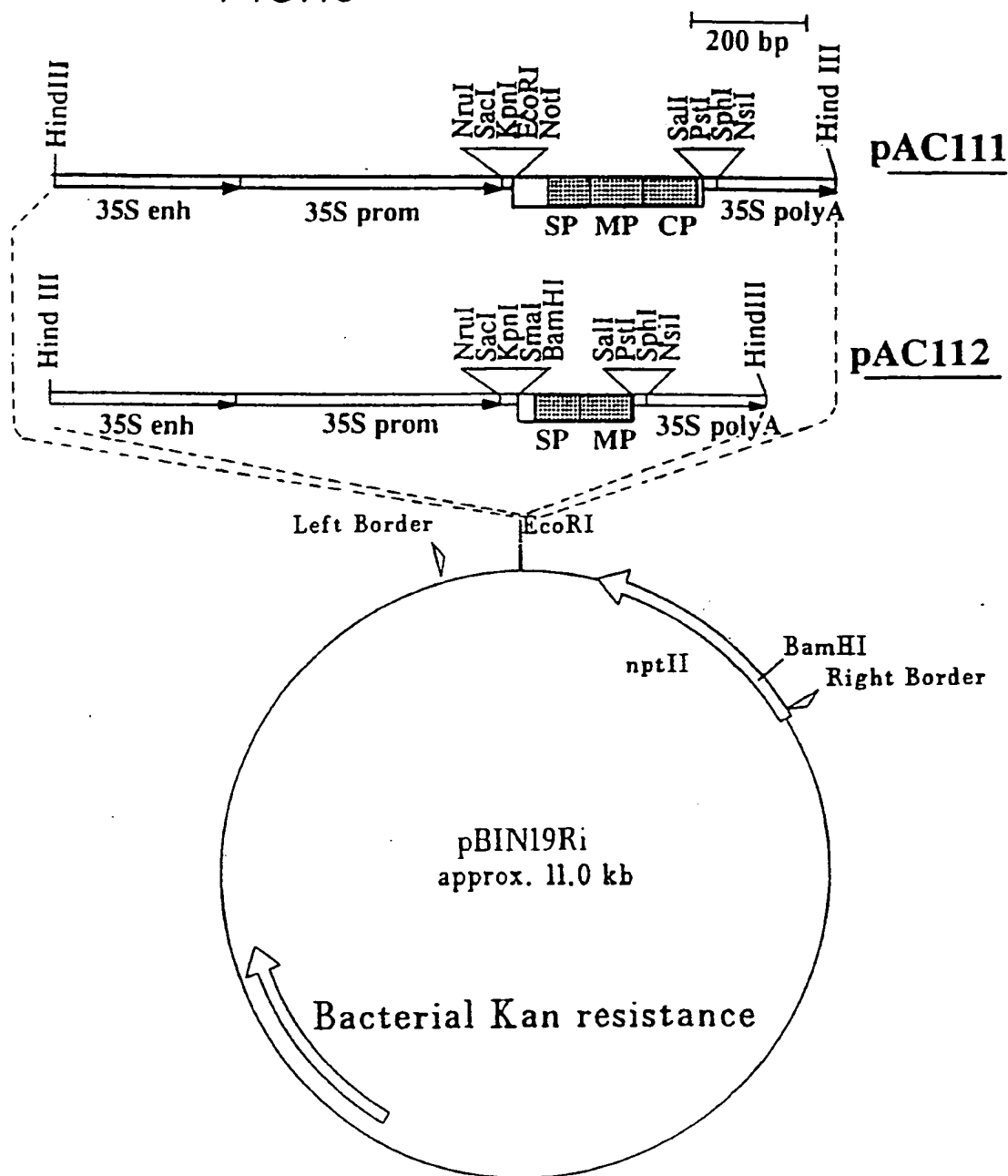
10 / 11

FIG. 9B



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FIG. 10



INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 92/00999

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) *		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC ⁵ : C 07 K 7/10, C 07 K 3/02, C 07 K 3/28, C 12 N 15/03, C 12 N 15/05, C 12 N 15/29, C 12 N 15/82, A 01 H 5/00, A 61 K 37/02		
II. FIELDS SEARCHED		
Minimum Documentation Searched *		
Classification System	Classification Symbols	
IPC ⁵	C 07 K 3/00, C 07 K 7/00, C 07 K 13/00, C 12 N 15/00, A 61 K 37/00, A 01 H 5/00	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are included in the Fields Searched *		
III. DOCUMENTS CONSIDERED TO BE RELEVANT *		
Category *	Citation of Document, ** with indication, where appropriate, of the relevant passages **	Relevant to Claim No. **
P, X	Chemical Abstracts, vol. 116, no. 21, issued 1992, May 25 (Columbus, Ohio, U.S.A.), W.F. BROEKAERT et al. "Anti- microbial peptides from Amaranthus caudatus seeds with sequence homology to the cysteine/glycine-rich domain of chitin-binding proteins", see page 410, column 1, the abstract-no. 211 129a, Biochemistry 1992, 31(17), 4308-14 (Eng).	1-22
A	EP, A1, 0 388 186 (E.I. DU PONT DE NEMOURS) 19 September 1990 (19.09.90), see claims 6-8; page 92, lines 7-11; claim 50; page 94, lines 52-57.	11-14
<p>* Special categories of cited documents: **</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"Z" document member of the same patent family</p>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
22 October 1992	11 NOV 1992	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	SCHARF e.h.	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
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A	EP, A1, 0 354 687 (E.I. DU PONT DE NEMOURS) 14 February 1990 (14.02.90), see claims 3-7; page 28, lines 5-15. --	11-14
A	EP, A2, 0 334 383 (MAX-PLANCK-GESELLSCHAFT) 27 September 1989 (27.09.89), see claims 9-12; page 12, column 1, lines 42-58; column 2, lines 1-19. --	11-14
A	Chemical Abstracts, vol. 100, no 9 issued 1984, February 27 (Columbus, Ohio, U.S.A.), A. BATRA et al. "Gas chromatography identifi- cation of fatty acid methyl esters and antimicrobial activity of Amaranthus hybridus(Amaranthaceae) oil", see page 487, column 2, the abstract-no. 66 856y, Grasas Aceites (Seville) 1983, 34(5), 329-31 (Eng). --	17,18
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A	Chemical Abstracts, vol. 112, no. 5, issued 1990, January 29 (Columbus, Ohio, U.S.A.), S.J. RINDERLE et al. "Isolation and characteri- zation of amaranthin, a lectin present in the seeds of Amaran- thus caudatus, that recognizes the T- (or cryptic T-) antigen", see page 435, column 2, the abstract-no. 34 252e, J. Biol. Chem. 1989, 264(27), 16123-31 (Eng). ----	19-21

ANHANG

zum internationalen Recherchen-
bericht über die internationale
Patentanmeldung Nr.

ANNEX

to the International Search
Report to the International Patent
Application No.

ANNEXE

au rapport de recherche inter-
national relatif à la demande de brevet
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In diesem Anhang sind die Mitglieder
der Patentfamilien der in obenge-
nannten internationalen Recherchenbericht
angeführten Patentedokumente angegeben.
Diese Angaben dienen nur zur Unter-
richtung und erfolgen ohne Gewähr.

This Annex lists the patent family
members relating to the patent documents
cited in the above-mentioned inter-
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in no way liable for these particulars
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of information.

La présente annexe indique les
membres de la famille de brevets
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dans le rapport de recherche inter-
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ments fournis sont donnés à titre indica-
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